

REVIEW ARTICLE

Pheromones in Aphids: A Review

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ABSTRACT: Aphids release an alarm pheromone on being attacked by predators to signal other members of the colony to evacuate and to make sure they do not become preys to natural enemies. This pheromone induces the individuals to produce winged offsprings which can fly away from the plant to avoid predators. The most common constituent in the alarm pheromone was found to be $(E) - \beta$ – Farnesene. It is also found that these pheromones released by the Aphids can be a trail for attracting natural enemies towards colonies and hence causing self harm. As this pheromone can cause harm to the aphids themselves, it is of a great advantage to plants which produce $(E) - \beta$ – Farnesene as one of the volatiles suspected to be a naturally resistant against Aphid infestations. Plants releasing this volatile are found to be protecting their sap against these aphid infestations as these pests are able to perceive these volatiles and avoid sucking the sap and disperse to avoid being attacked by enemies.

Keywords: Aphids, pheromones, $(E) - \beta$ – farnesene, nepetalactol, nepetalactone

INTRODUCTION

Chemical signals and pheromones in particular play a very important role in agricultural pest management and help in improving the potential of various insecticides and drastically reduce the harmful effects on the environment (L. J. Wadhams. 1990). Aphids are polyphagous sucking pests which are capable of attacking various plants and causing severe damage. They suck the sap of the plants and produce toxic saliva over the plants leading to decrease in the plant's ability to conduct photosynthesis and also transmit plant viruses causing various diseases (Sarah H. Dewhirst et al., 2010). They also produce excreta in the form of Honeydew which leads to growth of moulds in the host plants (Jia Fan et al., 2015). Aphids are considered to be model organisms to study olfaction and chemical ecology. During higher temperatures they live in small groups and reproduce by parthenogenesis Mondor et al., 2000). When aphids are attacked by natural enemies they produce an alarm pheromone after which they display behavior of producing winged forms that leave the plant or they just walk away to another plant with better nutrition due to predator pressure. This sticky secretion is produced by a pair of cornicles present on the dorsal surface of their body. This prevents their predators and parasitoids from feeding on them. This is also in indication for other members of the colony to move away from the host and get away from danger (Michael H. et al., 2006). Some of the plants are also found to be producing volatiles having the same composition as that of the Aphid alarm pheromone which may be known to prevent these pests from settling on the plant and leads to a reduced nutrition intake by them which prevents their reproduction (Kunert et al., 2010). This pheromone not only acts as a signal between aphid individuals but also acts as an interacting medium between aphids and plants and aphids and their natural enemies (Vandermoten et al., 2012). On examination of various aphids species, studies suggest that 21 out of 23 species contain (E) – β – Farnesene in their pheromones (Huili Qiao *et al.*, 2009). The release of these alarm pheromone component may not only be beneficial to the aphids but also can be damaging because they will have to leave the host plant loosing out on food supply, increased risk of high mortality in individuals due to relocation and inviting predators to be attacked. To avoid these damages as much as possible the aphids may release the alarm pheromones only when physically attacked by the natural enemies and not when they detect predators indicating that they have a chance to preserve life and food (Christoph Joachim et al., 2013).

PLANT VOLATILES AGAINST APHID INFESTATIONS

Volatiles mediate interactions occurring between the plants, aphids and the external environment. Plants have to capacity to show defense against various pathogens and pests by the ability to change their gene expression and metabolism rate (Martin de Vos *et al.*, 2010). Volatiles released by plants help in protection from pest infestations like that of aphids. As the constituents in

the plant volatiles are found to be the same as the alarm pheromones produced by aphids, the plant is sought to be gaining a benefit from this due to a decrease in the aphid feeding and reproduction rates. This urges the aphids to produce more number of winged off springs which show behavior of leaving the host plant. This type of behaviour has been observed in Pea and Cotton species of aphids (Grit Kunert et al., 2010). Studies by (Grit Kunert et al., 2010) reveal that the continuous production of these volatiles from plants might not have a direct effect on these pests. It was found that some natural enemies of aphids may be attracted to these volatiles which may have a positive effect of controlling the infestations on aphid host plants. It is also well documented that plant volatiles like 2-Phenylethanol produced a response in the olfaction of aphid predators and parasitoids like lacewings which attract them towards the aphid host plants as a beneficiary act to get rid of infestations. Studying the olfactory responses between in the prey's host plant and the predators helps in development of efficient biological control in the field of agriculture to prevent invasive and harmful pests (Zhu et al., 2005). Pyrethrum flowers are found to be attracting aphid predators like ladybugs by producing (E) $-\beta$ - Farnesene indicating this pheromone helps predators find aphid preys and acts as a natural biological control. They exhibit defense mechanism from aphids by the production of these pheromones protecting the flowers which is the most important part of any plant (Jinjin Li et al., 2019). Some species of wild potato are found to be metabolizing (E) $-\beta$ – Farnesene as a volatile component due to which it acts as a natural resistance against aphid infestations (Huili Qiao et al., 2009). Aphid parasitoid behavior of how they are attracted to their prey by chemical cues can be studied by using volatiles released by plants like mimicking pheromones produced by aphids like (E) $-\beta$ - Farnesene (Alarm pheromone) and Nepetalactone (Sex Pheromones). Using these pheromones to attract these parasitoids can be used as a Bio-Control in Agriculture to stop aphid infestations on important economic plants (Ameixa et al., 2011). In nature, plants follow a 'push and pull theory' where the volatiles push the aphid pests away and pull the natural enemies towards themselves for the environmental benefit. This helps in the study of aphid to plant, aphid to aphid, aphid to natural enemies and natural enemies to plant interactions (Jia Fan et al., 2015).

PRODUCTION OF WINGED FORMS OF APHIDS

Studies demonstrated by (Grit Kunert *et al.*, 2005) show that when these aphids are exposed to artificial

alarm pheromones just like the ones they produce naturally, this resulted in production of winged aphids in groups which was referred to as 'crowding'. Wild potato has found to be producing (E) $-\beta$ - Farnesene which acts as a natural repellent against aphids. (Michael H. et al., 2006) suggests that this phenomenon can be used to as a technique of crop protection from aphids by genetically modifying the crop to produce (E) $-\beta$ – Farnesene. On exposure to (E) – β – Farnesene under laboratory conditions aphids were found to be producing good number of winged off-springs, but studies by (Hatano et al., 2010) suggest that there was also a high proportion of winged individuals produced under field conditions. Sometimes production of winged morphs are also induced by the females if they are under some sort of stress like overcrowding of individuals on the host or non-availability of food on the host plant. In the case of Pea aphids, it was found that there is an increased production of winged individuals in large colonies due to more of physical contact among them which triggers the competition for food and space. Winged morphs are produced less in number in smaller colonies due to lower physical contact among individuals (Hatano et al., 2010). Aphids in which the antennae were removed did not perceive (E) – β – Farnesene component of the alarm pheromone and did not produce wings to leave the host plant indicates that the pheromone signals could be perceived only by the olfactory system (Jia Fan et al., 2015). Winged forms of Aphids also termed as 'alate' forms are not considered to be strong fliers but they can manage to fly to a good distance by drifting along with the wind. As aphids are pests that transmit plant viruses, some of them which are dependant on insect vectors for transmission can rely on these 'alate' forms of aphids which not only transmit diseases to other parts for the plant but also to different plants (Martin de Vos et al., 2010).

A MUTUALISTIC RELATIONSHIP BETWEEN APHIDS AND ANTS

Aphids and ants are found to be engaging in a relationship of mutualism as ants get a continuous supply of honey dew, an excretion produced by aphids which is rich in carbohydrates and water and functions to be their good source of nutrition whereas in return aphids acquire protection and sanitation from the ants as they clean up all the trails of honey dew. Aphids show behavior of dispersal from the host plants by forming winged individuals when attacked by predators, but this may lead to loss of food supply to the ant colony. Hence the ants come up with various strategies to decrease their

dispersal. These various strategies may include applying hormones on the aphids nymphs to prevent wing formation and to prevent them from flying away. The fire ants apply trial pheromones along the route that they take which leads them to the food source and this trail is maintained until the trail leading to the food source is not available anymore (Tian Xu et al., 2021). (Verheggen et al., 2012) suggested that ants have shown orientation and attraction towards the pheromones released by aphids which has been proved with olfactometer and choice assays while other volatiles did not show any response. Ants in association with aphids show behavioral responses on the abdomen of the aphids to generate increased release of honeydew and on exposure to the components present in the alarm pheromone of the aphids they prepare themselves to attack aphid predators as they provide protection to them. Ants are found to be locating their aphid partners for food by using low levels of the alarm pheromones produced by aphids when they are not attacked by predators. When aphids are attacked they tend to produce high amounts of alarm pheromone which urge the ants to show aggressive behavior intending to kill the aphids predators. On application of (E) – β – Farnesene, ants in mutualistic relationship with aphids are seen to be becoming very aggressive and attacking the aphid predators to protect them (Eduardo Hatano et al., 2010). When aphids are attacked by predators they constantly release the alarm pheromone containing (E) $-\beta$ - Farnesene which is perceived by the neighboring aphids in the colony and they disperse from the colony immediately as a result of threat but when the colony is attended by ants, the aphids do not disperse as they exhibit a mutual relationship and depend on the ants for protection (Acar et al., 2001).

SEX PHEROMONES AND OTHER PHEROMONES

Mature individuals on the colonies release sex pheromones which attract mates of conspecific species for sexual reproduction. Other pheromones they release are found to be called Aggregation pheromone. This is released when a single individual has landed on a new host plants and signals others indicating them about food availability (Dewhirst *et al.*, 2010). Aphid females produce sex pheromones to attract sexually mature males but predators and parasitoids like lacewings adults are also found to be getting attracted by these sex pheromones called Nepetalactol. Many aphid species belonging to the Family Aphidae produce sex pheromones that consist of a mixture of Nepetalactol and Nepetalactone belonging to the category of Iridoids (Cyclopentanoids) (Dewhirst *et al.*, 2008). Sexual female aphids produce Nepetalactol in their sex hormones which also function as Aggregation hormones which aid in accumulation of large population to ease the process of mating. This not only helps in females finding male mates but also allows them to look for various locations to produce their sexual female progeny (Park et al., 2000). It is also observed that during summer and autumn season oculata species of lacewings are found to be attacking Soyabean aphids and are specifically located in the regions of their prey presence indicating that aphid pheromones are having a major role to play in attracting predators (Zhu et al., 2005). Pheromones produced by herbivorous insects are detected by their various natural enemies and assist in prev localization and can be termed under the category of Kairomones (Christoph Joachim et al., 2015). P. humuli species of male aphids are found to be responding to sex pheromones from a distance of three meters indicating they can locate their mates from a long distance. Electroantennogram studies reveal that the primary Rhinaria in aphids (hairless region at the tip of the snout) detect a number of volatiles and sex pheromones at different positions of the antennae. In contrast to these studies the secondary Rhinaria are found to be detecting only sex pheromones in sexually mature females (Gynoparae) and males (Tom et al., 2004).

Alarm pheromones produced by aphids having (E) $-\beta$ - Farnesene can have both positive and negative effects. The production in of this in an overcrowded colony can help in dispersal of individuals but the constant release of this can invite trouble to aphids by attracting predators and natural enemies. Studies show that female hoverflies being one of the major predators for aphids lay eggs not depending upon the number of prey in the colonies but completely depend on the amount of evoking pheromone substances produced by the aphids and also the concentration of volatiles produced by the host plants (Almohamad et al., 2008). (Verheggen et al., 2008) demonstrates that the alarm pheromone produced by aphids can not only be trigger predator attacks but can also be released and accumulated in response to some odor signals released by other colony members in aphid juveniles during their developmental stages. When a colony experiences a potential threat by certain predator aphids they may have to adapt to such contagious responses as a small amount of pheromone released by very few aphids may not be enough to warn the entire colony. However when the Acyrthosiphon pisum species of aphids were exposed to alarm pheromone signals they showed a typical behavior of moving away and the results showed that they did not produce the excess of pheromone to warn the other colonies indicating that the contagious responses of (E) – β – Farnesene does not occur (Verheggen *et al.*, 2008). Lures containing aphid sex pheromone components like Nepetalactol and Nepetalactone were used for field evaluation to check for aphid abundance. Results indicated that though there was a variance in the aphid population there was a significant decrease their abundance due to attraction of parasitoids and natural predators causing a drop in their numbers. Thus utilizing the semio-chemicals derived from the host or prey can function as a good natural bio-control in avoiding pests on crops in field conditions (Yoshitaka Nakashima *et al.*, 2016).

Studies conducted by (Boo et al., 2003) in the fields of Korea revealed that the lacewing species Chrysopa cognata being a major aphid natural predator was highly attracted to the aphid sex pheromone component Nepetalactol than the other component Nepetalactone. When both the component chemicals were mixed to check for responses, the number of Chrysopa cognata increased in the trap along with the increase in the concentration of Nepetalactol but the number of was never found to be higher than the count achieved when Nepetalactol was used alone (Boo et al., 2003). Vials containing Aphid sex pheromone component was used as a technique to check for attracting aphid parasitoids in field conditions. These vials containing the pheromone components were placed at different distances from the aphid infested plants to analyze at what particular distance the parasitoids would get attracted to aphids. Traps placed next the infested plants or at a short distance from it showed a significant increase in the attacks on aphid colonies whereas the traps placed at a far distance showed no effect on the parasitoid attraction indicating that they could not perceive the pheromones at very long distances. Observations indicated that the placement of these vials having the pheromone components increased the attacks on aphids by natural enemies. These experiments gives a proof that the natural enemies of aphids are not just attracted by the pheromone trap but are found to be actively attacking the aphids around the spaces of the trap to mummify them (Glinwood et al., 1998). Other than the major component (E) $-\beta$ - Farnesene which is a sesquiterpene in the aphid alarm pheromone there are also many other components like Monoterpenes and analogs which have not been investigated as much as (E) $-\beta$ – Farnesene. It was discovered that the combination of all these multiple components were attracting a large number of predators when compared to only a single component of (E) – β – Farnesene. The attractiveness of these predators also increased with increase in the concentrations of the multiple components (Yaoguo Qin et. al., 2022).

The adult aphids emit a lower concentration of $(E) - \beta$ – Farnesene in their pheromone droplets when compared to the juveniles or pre-reproductive aphids. Reasons for this can be stated as pre-reproductive aphids are immature and can invest most of their metabolism into producing the alarm pheromone as they have low

In recent times, efforts have been made to conserve natural bio-control processes which involves enhancing the natural populations of aphid natural enemies to control infestations on economically important agricultural and horticultural crops. For locating their hosts, aphid natural enemies use the chemical information produced by their host. Infested plants with aphids were found to be inducing the neighboring un-infested plants to produce the same volatiles containing pheromones which could attract natural enemies. This was a behaviour shown by the infested plant to increase the signal potential to attract more predators. This behavior was physiologically induced by the infested plants when the rhizosphere of both the plants came in contact with each other when grown together in a same pot and the roots came in contact with each other (Wilf Powell et al., 2003). Studies indicate that the attraction experienced by predators to the pheromones released by the hosts are not always uniform. The concentrations released completely depend on the type, intensity and the species that are involved in the attack. In the case of aphids, natural predators like ladybugs were found to be attracted to smaller concentrations of (E) $-\beta$ – Farnesene and attacked the hosts very quickly when compared to lacewings which required higher concentrations (Joachim et al., 2013). Lady beetles are considered to be one of the key factors used as bio-control to protect major crops from aphid infestations and also various other pests. Studies suggest that they can perceive (E) $-\beta$ – Farnesene produced in the aphid alarm pheromone indicating that they have evolved along with a strong sensory system which can detect odors released by their prey. Lady beetles respond to various odors like these when they are in need of food, shelter and a host to reproduce. It is said that Lady beetles adults and larvae not only respond to olfactory signals but can also locate their host prev by recognising colours (EB Acar et al., 2001). It was found that juvenile aphids tend to release large amounts of (E) $-\beta$ - Farnesene in their alarm pheromones when compared to reproductively active adults as they reside in large clusters of colonies and are more likely to be predated when adults tend to move away from the host to different places. As the aphids develop there is surely in increase in the size of the droplets released by their cornicles but there is no increase in the concentration of (E) $-\beta$ – Farnesene.

reproductive values. Another reason can be that younger aphids can be very easily killed by predators and natural enemies whereas adults are capable of leaving the host quickly by flying away or dropping off (Mondor et al., 2000). Sometimes the amount of alarm pheromone emitted by the aphids may not be enough to alert all the individuals as the odor may get diluted along with the blowing wind. To overcome this the un-attacked aphids may also release more alarm pheromone in response to the signals received by the attacked individuals to double the amount and to also alert a large number of individuals from threat. This phenomenon still remains to be unknown as to whether they show such behavior or not. But amplification of the pheromones, instead of being beneficial may also turn out to be risky as they may invite more number of predators to the colony due to strong and concentrated emission (Eduardo Hatano et al., 2008). Orchids come under the category of deceptive flowers which get the service of pollination by the pollinators but do not give the pollinators anything in return for the service provided. They often get their services done by the pollinators by deceiving them either by visual, olfactory or both.

The terrestrial orchid species Epipactis veratrifolia emit fragrances which mimic the alarm pheromones produced by aphids attracting aphid natural enemies like hoverflies for pollination services. These hoverflies are considered to be beneficiary insects to the orchid flowers and show oviposition behavior of laying eggs which is also derived from the aphid produced kairomones. (E) $-\beta$ - Farnesene is considered to be one of the major components of the aphid alarm pheromone but there are also other terpenoids which also conduct the action of alarm pheromones. A very common species of aphids in the Middle East called M.Viciae produces α - and β -pinene, and β -myrcene as a component in their alarm pheromones. The odor of these components were found to be very similar to the odor produced by the orchid flowers Epipactis veratrifolia. Studies suggest that orchid flowers do not mimic one particular species of aphid pheromones as they do not produce the exact amount and composition of pheromones produced by a specific species. This type of mimicry by the orchid flowers are completely justified as all the hoverfly species do not feed on one particular species of aphids (Johannes Stokl et al., 2010). The pheromones produced in insects are considered to be in very small and trace amounts making it difficult to identify them, as they are also associated with various other substances. Gas Chromatography (GC) serves to be a highly efficient technique in the identification of various complex and natural substances but other additional techniques are also required along with it to isolate the biological products in the component. Small fractions of biological material obtained from the Gas Chromatography has been used for assay experiments to test the activity of insect antennae in the Electro-antennogram (EAG) (Wadhams et al., 1990). As a defense mechanism Aphids have also shown a behavior of smearing the droplets released by their cornicles containing the alarm pheromone along the body of the predator which alerts the colony members of an approaching predator (Ezra G. Schwartzberg et al., 2008). Aphids when reared in isolation under laboratory conditions produced less amount of alarm pheromones when compared to aphids reared along with other members. This behavior indicates that they prefer social environments and utilize the signals given by their neighbors to stimulate the pheromone production (Verheggen et al., 2009). Genetically modifying plants with genes to produce volatiles similar to the Aphid alarm pheromone (E) $-\beta$ - Farnesene is gaining importance in recent times as a good biocontrol agent to protect plants from infestations. This process has been successfully achieved by developing the hexaploid wheat cv. Cadenza. The volatiles released by this crop has shown successful results by preventing aphids from infesting and also attracts aphid natural enemies by the chemical odor (Bruce et al., 2015).

ODORANT BINDING PROTEINS

The peripheral nervous system of insects consist of Odorant binding proteins (OBP) which regulate the antenna to detect various olfactory and gustatory senses (Qian Wang et al., 2021). OBP help in perceiving different odors and aids in their discrimination (Huili Qiao et al., 2009). They conduct chemo-sensation and chemical signals to identify volatiles on their host plant and also to locate their mates using sex pheromones (Qian Wang et al., 2021). The hydrophobic volatile compounds in the environment are proposed to be transmitted to the insect olfactory system through these OBPs (Jia Fan et al., 2017). OBPs transport the odor fragrances through the sensory neurons of the insect olfactory system. Their sensory receptors contain proteins of high concentrations but do not interact with each other in localized areas. Knocking down the genes involved in the OBP production showed that the Aphids were still being repelled by the production of (E) – β – Farnesene (Qian Wang et al., 2021).

Among insects aphids specifically use their receptors in the primary and secondary Rhinaria present in the antennae to detect odors (Tom W. Pope *et al.*, 2004).

Studies indicate that OBP3 and OBP7 may be involved in the aphid olfaction for perceiving (E) – β – Farnesene in the alarm pheromone. Rhopalosiphum padi species of aphids were used to study the responses of olfaction for (E) $-\beta$ - Farnesene and volatiles extracted from crushed aphids. Results indicated that Rhopalosiphum padi was repelled by both (E) – β – Farnesene and the volatiles extracted from the crushed aphids (Jia Fan et al., 2017). Odorant receptors (ORs) are one of the sensitive chemosensory systems identified in aphids aiding in the perception of pheromones (Ruibin Zhang et al., 2017). ORs are basically complexes that are present in the insect olfactory system consisting of a large number of odor specific receptors and odor specific co-receptors (Orco) (Fan et al., 2015). An odorant receptor called ApisOR5 found to be expressing in the antennal segment of the aphids shows response to (E) – β – Farnesene when expressed along with Orca, an odorant receptor co-receptor. ApisOR5 is one of the receptors belonging to the large subfamily of odorant receptors (Zhang et al., 2017).

REFERENCES

- Acar, E. B., Medina, J. C., Lee, M. L. and Booth, G. M. 2001. Olfactory behavior of convergent lady beetles (Coleoptera: Coccinellidae) to alarm pheromone of green peach aphid (Hemiptera: Aphididae). *The Canadian Entomologist*, **133**(3): 389-397.
- Almohamad, R., Verheggen, F. J., Francis, F., Lognay, G. and Haubruge, E. 2008. Emission of alarm pheromone by non- preyed aphid colonies. *Journal* of *Applied Entomology*, **132**(8): 601-604.
- Ameixa, O. M. C. C. and P. Kindlmann. 2011. Effect of synthetic and plant-extracted aphid pheromones on the behaviour of *Aphidius colemani*. *Journal of Applied Entomology*, 136: 292-301.
- Beale, M. H., Birkett, M. A., Bruce, T. J., Chamberlain, K., Field, L. M., Huttly, A. K., Martin, J. L., Parker, R., Phillips, A. L., Pickett, J. A. and Prosser, I. M. 2006. Aphid alarm pheromone produced by transgenic plants affects aphid and parasitoid behavior. *Proceedings of the National Academy of Sciences*, **103**(27):10509-10513.
- Boo, K. S., Choi, M. Y., Chung, I. B., Eastop, V. F., Pickett, J. A., Wadhams, L. J. and Woodcock, C. M. 2000. Sex pheromone of the peach aphid, Tubero cephalus momonis and optimal blends for trapping males and females in the field. *Journal of chemical ecology*, **26**: 601-609.

- Boo.S. S., Kang, J. H., Park, J. A., Pickett and Wadhams, L. J. 2003. Field Trapping of *Chrysopa cognata* (Neuroptera: Chrysopidae) with Aphid Sex Pheromone Components in Korea. *Journal of Asia Pacific Entomology*, 6(1):29-36.
- Bruce, T., Gudbjorg, I., Aradottir, Lesley, E., Smart, Janet L., Martin., John C. Caulfield., Angela Doherty., Caroline A. Sparks., Christine M. Woodcock., Michael, A., Birkett, Johnathan A., Napier, Huw D., Jones and John A. Pickett. 2015. The first crop plant genetically engineered to release an insect pheromone for defence, *Scientific Reports*, 5: (11183).
- Christoph Joachim., Eduardo Hatano and Anja David., Maritta Kunert and Cornelia Linse and Wolfgang .W. Weisser. 2013. Modulation of Aphid Alarm PheromoneEmissionofPeaAphidPreybyPredators. *Journal of Chemical Ecology*, **39**:773-782.
- Dewhirst, S. H., John, A., Pickett and Jim Hardie. 2010. Chapter Twenty Two – Aphid Pheromones. *Vitamins and Hormones*, **83**:551-574.
- Dewhirst, S. H., Michael, A., Birkett., Jean .D., Fitzgerald, Alex Stewart-Jones., Lester, J., Wadhams., Christine M. Woodcock., Jim Hardie and John, A., Pickett. 2008. Dolichodial: A New Aphid Sex Pheromone Component?. Journal of Chemical ecology, 34:1575-1583.
- Fan, J. Yong Zhang., Frederic Francis., Dengfa Cheng., Jingrun Sun and Julian Chen. 2015. Orco mediates olfactory behaviors and winged morph differentiation induced by alarm pheromone in the grain aphid, Sitobion avenae. Insect Biochemistry and Molecular Biology, 64:16-24.
- Foster, S. P., Denholm, I., Thompson, R., Poppy, G. M. and Powell. W. 2005. Reduced response of insecticide-resistant aphids and attraction of parasitoids to aphid alarm pheromone; a potential fitness trade-off. *Bulletin of Entomological Research*, 95: 37-46.
- Glinwood, R. T., Powell, W. and Tripathi. C. P. M. 1998. Increased Parasitization of Aphids on Trap Plants Alongside Vials Releasing Synthetic Aphid Sex Pheromone and Effective Range of the Pheromone. *Biocontrol Science and Technology*, 8(4): 607-614.
- Hartfield, C. M., Campbell, C. A. M., Hardie, J., Pickett, J. A. and Wadhams, L. J. 2001. Pheromone traps for the dissemination of an entomopathogen by the damson-hop aphid *Phorodon humuli. Biocontrol Science and Technology*, **11**(3): 401-410.

- Hatano, E., Kunert, G. and Weisser, W.W. 2010. Aphid wing induction and ecological costs of alarm pheromone emission under field conditions. *PLoS One*, **5**(6): 11188.
- Hatano., Grit Kunert and Stefan Bartram., Wilhelm Boland and Jonathan Gershenzon and Wolfgang .W. Weisser. 2008. Do Aphid Colonies Amplify their Emission of Alarm Pheromone? *Journal of Chemical Ecology*, 34:1149-1152.
- Huili Qiao., Elena Tuccori., Xiaoli He., Angelo Gazzano., Linda Field., Jing-Jiang Zhou and Paolo Pelosi.
 2009. Discrimination of alarm pheromone (E)β-farnesene by aphid odorant-binding proteins. *Insect Biochemistry and Molecular Biology*, 39 (5-6): 414-419.
- Jia Fan., Wenxin Xue., Hongxia Duan., Xin Jiang., Yong Zhang., Wenjuan Yu., Shanshan Jiang., Jingrun Sun and Julian Chen. 2017. Identification of an intraspecific alarm pheromone and two conserved odorant-binding proteins associated with (E)-βfarnesene perception in aphid *Rhopalosiphum padi. Journal of Insect Physiology*, **101**:151-160.
- Jinjin Li., Hao Hu., Jing Mao., Lu Yu., Geert Stoopen., Manqun Wang., Rol and Mumm, Norbert C. A. de Ruijter., Marcel Dicke., Maarten A., Jongsma and Caiyun Wang. 2019. Defense of pyrethrum flowers: repelling herbivores and recruiting carnivores by producing aphid alarm pheromone. *New Phytologist*, **223**(3):1607-1620.
- Joachim, C. and Weisser, W.W. 2015. Does the aphid alarm pheromone (E)-β-farnesene act as a kairomone under field conditions?. *Journal of Chemical Ecology*, **41**: 267-275.
- Johannes Stokl., Jennifer Brodmann., Amots Dafni., Manfred Ayasse and Bill S. Hansson. 2010. Smells like aphids: orchid flowers mimic aphid alarm pheromones to attract hoverflies for pollination. *Proceedings of the Royal Society B: Biological Sciences.*
- John A. Pickett., Rudolf K. Allemannb and Michael A. Birkett. 2013. The semiochemistry of aphids. *Natural Product Reports*, **30**(10): 1277–1283.
- Junwei Zhu., Aijun Zhang., Kye-Chung Park., Tom Baker., Brian Lang., Russell jurenka., John J. Obrycki., William R. Graves., J. A. Pickett., D. Smiley., Kamlesh R. Chauhan and Jerome A. Klun. 2006. Sex Pheromone of the Soybean Aphid, *Aphis glycines* Matsumura and Its Potential Use

in Semiochemical-Based Control. *Environmental Entomology*, **35**(2): 249-257.

- Kunert, G., Reinhold, C. and Gershenzon, J. 2010. Constitutive emission of the aphid alarm pheromone, (E)- β -farnesene, from plants does not serve as a direct defense against aphids. *BMC Ecology*, **10**: 1-12.
- Kunert., Susanne Otto., Ursula. S. R., Rose., Jonathan Gershenzon and Wolfgang W. Weisser. 2005. Alarm pheromone mediates winged dispersal morphs in aphids. *Ecology Letters*, 8(6):596-603.
- Lewis, M. J., Prosser, I. M., Mohib, A. and L. M. Field. 2008. Cloning and characterization of a prenyltransferase from the aphid *Myzus persicae* with potential involvement in alarm pheromone biosynthesis. *Insect Molecular Biology*, **17**(4):437-443.
- Martin de Vos and Georg Jander. 2010. Volatile communication in plant–aphid interactions. *Current Opinion in Plant Biology*, **13**(4):366-371.
- Mondor, E. B., Baird, D. S., Slessor, K. N. and Roitberg, B. D. 2000. Ontogeny of alarm pheromone secretion in pea aphid, *Acyrthosiphon pisum. Journal of Chemical Ecology*, 26: 2875-2882.
- Park, K. C., Elias, D., Donato, B. and Hardie, J. 2000. Electroantennogram and behavioural responses of different forms of the bird cherry-oat aphid, *Rhopalosiphum padi*, to sex pheromone and a plant volatile. *Journal of Insect Physiology*, **46**(4):597-604.
- Qian Wang., Jing-Tao Liu., Yong-Jun Zhang., Ju-Lian Chen., Xian-Chun Li., Pei Liang., Xi-Wu Gao., Jing-Jiang Zhou and Shao-Hua Gu. 2021. Coordinative mediation of the response to alarm pheromones by three odorant binding proteins in the green peach aphid Myzus persicae. *Insect Biochemistry and Molecular Biology*, 130.
- Schwartzberg, E. G., Kunert, G., Stephan, C., David, A., Rose, U. S., Gershenzon, J., Boland, W. and Weisser, W.W. 2008. Real-time analysis of alarm pheromone emission by the pea aphid (Acyrthosiphon pisum) under predation. Journal of chemical ecology, 34:76-81.
- Stewart-Jones, A., Dewhirst, S.Y., Durrant, L., Fitzgerald, J. D., Hardie, J., Hooper, A M., Pickett, J. A. and Poppy, G. M. 2007. Structure, ratios and patterns of release in the sex pheromone of an aphid, *Dysaphis plantaginea. Journal of Experimental Biology*, 210(24): 4335-4344.

- Tian Xu., Meng Xu., Yongyue Lu., Wenqian Zhang., Jianghua Sun., Rensen Zeng., Ted C. J., Turlings and Li Chen. 2021. A trial pheromone mediates the mutualism between ants and aphids. *Current Biology*, **31**(21):4738-4747.
- Tom, W., Pope., Colin, A. M., Campbell., Jim Hardie and Lester, J. Wadhams. 2004. Electroantennogram responses of the three migratory forms of the damson-hop aphid, *Phorodon humuli*, to aphid pheromones and plant volatiles. *Journal of Insect Physiology*, **50**(11):1083-1092.
- Vandermoten, S., Mark, C., Mescher., Frederic Francis., Eric Haubruge and Francois, J. Verheggen. 2012. Aphid alarm pheromone: An overview of current knowledge on biosynthesis and functions. *Insect Biochemistry and Molecular Biology*, 42(3):155-163.
- Varnika Bhatia., Jaya Maisnam., Ajay Jain., Krishan Kumar Sharma and Ramcharan Bhattacharya. 2015. Aphid-repellent pheromone E-β-farnesene is generated in transgenic *Arabidopsis thaliana* over-expressing farnesyl diphosphate synthase 2. *Annals of Botany*, **115**(4): 581-591.
- Verheggen, F. J., Haubruge, E., De Moraes, C. M. and Mescher, M. C. 2009. Social environment influences aphid production of alarm pheromone. *Behavioral Ecology*, **20**(2): 283-288.
- Verheggen, F. J., Mescher, M. C., Haubruge, E., Moraes, C. D. and Schwartzberg, E. G. 2008. Emission of alarm pheromone in aphids: a noncontagious phenomenon. *Journal of Chemical Ecology*, 34:1146-1148.
- Verheggen., Lise Diez., Ludovic Sablon., Christophe Fischer., Stefan Bartram., Eric Haubruge and Claire Detrain. 2012. Aphid Alarm Pheromone as a Cue for Ants to locate Aphid Partners. *PLOS ONE*, 7(8).

- Wadhams, L. J. 1990. The use of coupled gaschromatography: electrophysiological techniques in the identification of insect pheromones. *Chromatography and Isolation* of Insect Hormones and Pheromones, 289-298.
- Wilf Powell and John A Pickett. 2003. Manipulation of parasitoids for aphid pest management: progress and prospects. *Pest Management Science*, 59:149-155.
- Yaoguo Qin., Shangyang Zhang and Zhengxi Li. 2022. Kairomonal Effect of Aphid Alarm Pheromones and Analogs on the Parasitoid *Diaeretiella rapae*. *Insects*, **13** (1055).
- Yoshitaka Nakashima., Takashi Y. Ida., Wilf Powell., John A. Pickett., Michael A. Birkett., Hisatomo Taki and Junji Takabayashi. 2016. Field evaluation of synthetic aphid sex pheromone in enhancing suppression of aphid abundance by their natural enemies. *Bio Control*, 61 (5):485-496.
- Yu Feng Sun., Filomena De Biasio., Hui Li Qiao., Immacolata Iovinella., Shao Xiang Yang., Yun Ling., Lea Riviello., Donatella Battaglia., Patrizia Falabella., Xin Ling Yang and Paolo Pelosi. 2012. Two Odorant-Binding Proteins Mediate the Behavioural Response of Aphids to the Alarm Pheromone (*E*)-β-farnesene and Structural Analogues. *PLOS ONE*, 7(3).
- Zhang, R., Bing Wang., Gerarda Grossi., Patrizia Falabella., Yang Liu., Shanchun Yan., Jian Lu., Jinghui Xi and Guirong Wang. 2017. Molecular Basis of Alarm Pheromone Detection in Aphids. *Current Biology*, 27(1):55-61.
- Zhu, J., Obrycki, J. J., Ochieng, S. A., Baker, T. C., Pickett, J. A. and Smiley, D. 2005. Attraction of two lacewing species to volatiles produced by host plants and aphid prey. *Naturwissenschaften*, **92**: 277-281.

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