Anti-insect activities of solvent extracts of *Andrographis paniculata* on the Diamondback moth, *Plutella xylostella* (L.)

R. SANGAVI^{1*}, Y. S. JOHNSON THANGARAJ EDWARD², S. JEYARAJAN NELSON³, M. PARAMASIVAM⁴, and C. MEENAMBIGAI⁵

^{1*}Ph. D. Scholar, Department of Agricultural Entomology, Navsari Agricultural University, Navsari, Gujarat, India ^{2,3,4}Department of Agricultural Entomology, Tamil Nadu Agricultural University, Coimbatore, Tamil Nadu, India

⁵Ph. D Scholar, Department of Agricultural Entomology, Bidhan Chandra Krishi Viswavidyalaya, Nadia, West Bengal, India

ABSTRACT: Diamondback moth, *Plutella xylostella* (Linnaeus.) is a key pest on *Brassicaceae* crops causing severe yield loss worldwide. Insecticidal properties of the solvent extracts of *Andrographis paniculata* against diamondback moth were studied as an alternative to insecticides. Among the extracts tested, Cent percent mortality was observed in hexane extract of *A. paniculata* at 0.6 per cent concentration. Maximum of 65.00 and 90.00 per cent mortality was observed in ethanol and ethyl acetate extract at 0.6 per cent concentration respectively.

Keywords: Diamond back moth, plant extract, efficacy

INTRODUCTION

In India, a total of 37 insect pests have been reported to feed on cabbage, of which the Diamondback moth (DBM), Plutella xylostella (L.) causes the loss of about 35 per cent with intensive control measures (Mohan and Gujar, 2003). Estimated total costs associated with damage and management of DBM worldwide was 4-5 billion US dollers per annum (Zalucki et al., 2012) and in India, the estimated cost for the control of DBM is about 16million US\$ per annum. Utilization of conventional synthetic insecticides posed certain problems such as adverse effects on natural enemies, development of resistance in target pests and pest resurgence. Hazardous implications of these pesticides and their residue at various trophic levels have also caused incalculable damage to every aspect of environment, globally. In India, the first report of P. xylostella resistance to insecticides (DDT and parathion) was made by Verma and Sandhu (1968) in Ludhiana (Punjab). Some population of P. xylostella developed resistance to new generation insecticides such as spinosad, avermectins, indoxacarb, emamectin benzoate and Bacillus thuringiensis, B.t. Cry toxins in the field (Pu et al., 2010)

MATERIALS AND METHODS

Soxhlet extraction

The powdered leaf of *A. paniculata* (100g) was sequentially extracted with 700 ml of hexane (non polar), ethyl acetate (medium polar) and ethanol (high

polar) solvent on soxhlet's extraction apparatus for 24 - 72 hours according to the plants used. The solvents were evaporated in a rotary vacuum evaporator at 40°C. The obtained extracts were pale yellow to pale brown in colour, viscous liquid, having a pleasant woody and spicy odour.

Evaluation of Solvent Extracts

The leaf dipping bioassay method described by Tabashnik and Cushing (1987) [15] was adopted to evaluate the insecticidal action of solvent extract against *P. xylostella* larvae. Cabbage leaves were washed with distilled water and dried for about10 minutes. Solvent extract of *A. paniculata* were tested at three concentration *viz.*, 0.2, 0.4 and 0.6 per cent along with NSKE 5 per cent as standard check. Completely untreated leaves and ethyl acetate treated leaves were used as control.

Fresh cabbage leaf discs (7.5 cm diameter) were cut from fully expanded cabbage leaves. The discs were dipped for 60 seconds in the test solutions. These treated and air dried leaves were placed in a petriplates lined with moist filter paper. In each petriplate 10third instar larvae was released using a camel hair brush and allowed to feed for 48 hours. After 48 hours, treated leaves were removed and fresh untreated leaves were given. Three replications of each of the treatment with 10 larvae per replicate were maintained. Readings were taken at 24 hours intervals up to adult emergence for larval mortality, pupal mortality and adult emergence. Any malformation in the treatments was also observed.

^{*} E-mail: sangavi.sabi@gmail.com

Statistical analysis

The data collected in the experiments were analysed by Completely Randomized Block Design (Gomez and Gomez, 1985). The data on per cent values and numbers were transformed into arcsine and square root values, respectively before subjecting them to statistical analysis.

RESULT AND DISCUSSION

Effect of hexane extract of A. paniculata on growth and development of P. xylostella

The mortality of P. xylostella when treated with different concentrations of hexane extract of A. paniculata is furnished in Table 1. On 1 DAT, the highest mortality (55.00 %) was recorded at 0.6 per cent, while 0.2 per cent concentration of A.paniculata caused only 5.00 per cent mortality. On 3 DAT, the mortality of P. xylostella due to A.paniculata, was cent per cent at 0.6 per cent while at 0.4 and 0.2 per cent concentrations, the mortality increased gradually as 40.00 and 50.00 per cent, respectively. After 5 DAT, more than 70.00 per cent mortality was recorded in all the concentrations of A. paniculata which was higher than hexane control and NSKE (60.00 and 30.00%, respectively) while the untreated check showed no mortality. The pupal mortality observed at 0.2 and 0.4 percent concentrations of hexane extract of A. paniculata was 5.00 and 20.00 per cent respectively. The least adult emergence was noticed in 0.6 and 0.4 per cent of A. paniculata (0.00 and 10.00 % respectively), while highest in NSKE (70.00 %).

Effect of ethanol extract of A. paniculata on growth and development of P. xylostella

On 1 DAT, no mortality was observed in all the three concentrations of ethanol extract of *A. paniculata*. However, on 2 DAT, 15.00 per cent mortality was observed at 0.2 and 0.6 per cent concentrations of *A. paniculata*. On 5 DAT, 70.00 per cent mortality was recorded at 0.2 per cent, while the mortality at 0.4 and 0.6 per cent concentrations increased gradually to 60.00

and 65.00 per cent, respectively. The pupal mortality was recorded in 0.4 and 0.6 per cent concentrations (20.00 and 15.00 % respectively). The normal adult emergence was recorded in the ethanol check which was found to be on par with untreated check (100 %) followed by NSKE at 5 per cent (50.00 %). Least adult emergence was observed at 0.4 and 0.6 per cent (20.00 %) which was on par with each other (Table 2).

Effect of ethyl acetate extract of A. paniculataon growth and development of P. xylostella

The mortality of *P. xylostella* when treated with different concentrations of ethylacetate extract of *A. paniculata* is furnished in Table 3. On 1 DAT, 70 per cent mortality was recorded at 0.6 per cent followed by 0.2 and 0.4 per cent (40.00 and 50.00 %respectively). On 5 DAT, 95 per cent mortality was found in 0.4 per cent concentration of the *A. paniculata* followed by 0.6 and 0.2 per cent (90.00 and 65.00 % respectively) treatments while the treated check showed 70.00 percent mortality and NSKE (40.00 %). The pupal mortality was observed in 0.2 and 0.6 per cent (5.00 %) which was found to be on par with each other. Least adult emergence was observed at 0.4 and 0.6 percent concentrations of ethyl acetate extract of *A. paniculata* (5.00 %) which were on par.

Similar result was reported by Govindarajan (2011), that methanol and ethyl acetate extracts of A. paniculata exerted cent per cent mortality at 200 ppm against C. quinque fasciatus and at 250 ppm against A. aegypti. Gautam et al. (2013) reported that flavonoid extract of flower buds of A. paniculata produced highest mortality (100%) at the concentration of 600 ppm for the late III or early IV instar larvae of A. aegypti and at the concentration of 200 ppm for the larvae of A. stephensi. Bright et al. (2001) reported that methanol and ethyl acetate extracts of A. paniculata at the concentrations (1000 ppm) lead to 72.01 and 67.69 per cent adult mortality of Callosobruchus chinensis. The alkaloid (14 deoxyandrographolide) present in A. paniculata which might be responsible for the larval mortality and antifeedant activity (Hermawan, 1994).

Table 1. Effect of hexane extract of A. paniculata on the growth and development of the diamondback moth, P. xylostella

Treatment		Lar	Larval mortality (%)	(0%)		Pupal mortality (%)	Adult	Adult emergence (%)
l	1 DAT	2 DAT	3 DAT	4 DAT	5 DAT		Normal	Malformed
Hexane extract (0.2%)	5.00 (12.92)°	15.00 (22.78) ^d	40.00 (39.22)°	75.00 (59.98) ^b	80.00 (63.41) ^b	5.00 (12.92) ^b	15.00 (22.78) ^d	1
Hexane extract (0.4%)	20.00 (26.55) ^b	50.00 (44.98) ^b	50.00 (44.98) ^b	70.00 (56.77)°	70.00 (56.77)°	20.00 $(26.55)^a$	10.00 (18.43)°	•
Hexane extract (0.6%)	55.00 (39.22) ^a	55.00 $(47.85)^a$	100.00 $(89.96)^a$	100.00 $(89.96)^a$	100.00 $(89.96)^a$	0.00 (4.05)°	0.00 (4.05) ^f	•
NSKE (5%)	20.00 (26.91) ^b	20.00 (26.91)°	25.00 (29.98) ^d	30.00 (33.19)°	30.00 (33.19)°	0.00 (4.05) [€]	70.00 (56.77) ^b	1
Hexane control	0.00 (4.05) ^d	0.00 (4.05)°	25.00 (29.99) ^d	60.00 (50.75) ^d	60.00 (50.75) ^d	0.00 (4.05)°	40.00 (39.22)°	•
Untreated control	0.00 (4.05) ^d	0.00 (4.05) ^e	0.00 (4.05)°	0.00 $(4.05)^{f}$	0.00 (4.05)f	0.00 (4.05) ^c	100.00 $(89.96)^{a}$	1
SE CD(0.05)	0.18	0.15	0.21	0.52	0.35	0.10	0.13	,

Table 2. Effect of ethanol extract of A. paniculata on the growth and development of the diamondback moth, P. xylostella

Treatment		Lar	Larval mortality (%)	(0%)		Pupal mortality (%)	Adul	Adult emergence (%)
I	1 DAT	2 DAT	3 DAT	4 DAT	5 DAT		Normal	Malformed
Ethanol extract (0.2%)	0.00 (4.05) ^b	15.00 (22.78) ^b	25.00 (29.99)°	60.00 (50.75) ^b	70.00	0.00 (4.05)°	30.00 (33.20)°	ı
Ethanol extract (0.4%)	0.00 (4.05) ^b	0.00 $(4.05)^{\circ}$	35.00 $(36.26)^{a}$	55.00 (47.85)°	60.00 (50.75)°	20.00 (26.55) ^a	20.00 (26.55) ^d	•
Ethanol extract (0.6%)	0.00 (4.05) ^b	15.00 $(22.78)^{b}$	30.00 (33.20) ^b	65.00 $(53.71)^a$	65.00 (53.71) ^b	15.00 (22.78) ^b	20.00 (26.55) ^d	ı
NSKE (5%)	15.00 (22.77) ^a	30.00 $(33.19)^a$	35.00 $(36.25)^{a}$	45.00 (42.11) ^d	50.00 (44.98) ^d	$0.00 \\ (4.05)^{\circ}$	50.00 (45.27) ^b	ı
Ethanol control	0.00 (4.05) ^b	0.00 $(4.05)^{\circ}$	0.00 $(4.05)^{d}$	0.00 (4.05)°	0.00 (4.05)°	$0.00 \\ (4.05)^{\circ}$	100.00 $(89.96)^a$	ı
Untreated control	0.00 (4.05) ^b	0.00 $(4.05)^{\circ}$	0.00 (4.05) ^d	0.00 $(4.05)^{f}$	0.00 (4.05)°	$\begin{array}{c} 0.00 \\ (4.05)^{\mathfrak{c}} \end{array}$	100.00 (89.96) ^a	ı
SE CD(0.05)	0.04	0.14	0.44	0.36	0.23	0.08	0.16	

Table 3. Effect of ethyl acetate extract of A. paniculata on the growth and development of the diamondback moth, Pxylostella

E		Larva	val mortality (%)	(%)		Pupal mortality(%)	Adult	Adult emergence(%)
	1 DAT	2 DAT	3 DAT	4 DAT	5 DAT		Normal	Malformed
Ethyl acetate extract (0.2%)	40.00 (22.78)°	45.00 (42.11)°	55.00 (47.85)°	75.00 (59.98) ^b	65.00 (53.71) ^d	5.00 (12.92) ^a	30.00 (33.20)°	
Ethyl acetate extract (0.4%)	50.00 (44.98) ^b	60.00 (50.75) ^b	65.00 (53.71) ^b	70.00 (56.77)°	95.00 (77.05) ^a	0.00 (4.05) ^b	5.00 (12.92) ^d	ı
Ethyl acetate extract (0.6%)	70.00 (56.77) ^a	70.00 (56.77) ^a	90.00 (71.54) ^a	90.00 (71.54) ^a	90.00 (71.54) ^b	5.00 $(12.92)^a$	5.00 (12.92) ^d	ı
NSKE (5%)	15.00 (22.77) ^d	30.00 (33.19)°	35.00 (36.25) ^d	40.00 (67.18) ^d	40.00 (39.21) ^e	0.00 (4.05) ^b	60.00 (50.74) ^b	ı
Ethyl acetate control	40.00 (22.78)°	40.00 (22.78) ^d	55.00 (47.85)°	70.00 (56.77)°	70.00 (56.77)°	0.00 (4.05) ^b	30.00 (33.20)°	ı
Untreated control	0.00 (4.05) ^e	0.00 $(4.05)^{f}$	0.00 (4.05) ^e	0.00 (4.05)°	0.00 (4.05) ^f	0.00 (4.05) ^b	100.00 (89.96) ^a	ı
SE CD(0.05)	0.37	0.16	0.65	1.25	1.25	0.04	0.12	

The present studies indicate that cent per cent mortality of P. xylostella larvae was observed in 5 DAT in 0.6 per cent concentration of hexane extract of A. paniculata and 85.00 and 40.00 per cent mortality was observed in 0.6 and 0.4 per cent concentrations of ethyl acetate and ethanol extracts, respectively.

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