



Escape resistance in fieldpea against *Phytomyza horticola* Goureau, as influenced by sowing time and its correlation with plant metabolites

RAJEEV KUMAR YADAV¹, ANUP CHANDRA^{2*}, ASHOK KUMAR PARIHAR², SUJAYANAND G.K.², VAIBHAV KUMAR², SANJAY M. BANDI², MEENAKSHI ARYA¹ and USHA¹

¹Rani Lakshmi Bai Central Agricultural University, Jhansi 284003, Uttar Pradesh

²ICAR-Indian Institute of Pulses Research, Kalyanpur, Kanpur 208024, Uttar Pradesh

*Email: anup.ento@gmail.com

ABSTRACT: Fieldpea or dry pea (*Pisum sativum* L.) is one of the important, highly productive cool season food legume crops and pea leaf miner, *Phytomyza horticola* Goureaui is a pest of high economic importance, associated with the crop. Twenty eight diverse field pea genotypes including checks were screened for their susceptibility to the pest and to study the influence of sowing time on the extent of infestation of pea leaf miner. The correlation of infestation with the host primary and secondary metabolites was also examined. In late sown condition, only eight genotypes were categorized as 'Susceptible', 2 genotypes under 'Moderately Resistant', 3 genotypes under 'Intermediate', whereas 15 genotypes were categorized as 'Moderately Susceptible'. None of the genotypes could be categorized as 'Very Highly Resistant', 'Highly Resistant', 'Resistant', 'Highly Susceptible' and 'Very Highly Susceptible' in case of late planting. In case of timely planting, only 9 genotypes were categorized under 'Moderately Susceptible', 7 genotypes each under 'Moderately Resistant' and 'Intermediate' and 5 genotypes grouped under 'Highly Resistant' category. None of the genotypes categorized as 'Susceptible' or beyond that in case of timely planting. Most of the genotypes which showed susceptibility under late sown condition had expressed more level of resistance in the timely sown crop. Although, different fieldpea genotypes differed significantly in terms of quantity of primary and secondary metabolites estimated, only the total phenol content was found significantly and positively correlated with the per cent infestation in the late sown genotypes.

Keywords: Genotypes, leaf miner, *Pisum sativum* L. resistance, screening, susceptibility

INTRODUCTION

Pea (*Pisum sativum* L.) is one of the oldest domesticated crops on the planet (Ambrose, 1995; Zohary and Hopf, 2000) as well as one of the important, highly productive cool season food legume crops grown around the world to consume as food, feed and fodder (Rubiales *et al.*, 2019). Since it is an excellent source of protein, starch, fibre and micro-nutrients, hence, widely used as an ingredient in many food industries around the world (Burstinet *et al.*, 2011; Dixit *et al.*, 2014; Gupta and Parihar, 2015; Parihar *et al.*, 2016). Seeds also contain vitamins, minerals, polyphenolics, galactosides, saponins, and phytic acid which are being studied for their health-promoting properties (Arnoldi *et al.*, 2015; Dahl *et al.*, 2012; Marles *et al.*, 2013; Mitchell *et al.*, 2009). It is a prominent pulse crop and an important part of sustainable cropping system (Duc *et al.*, 2010; Nemecek *et al.*, 2008).

Among the various biotic stresses associated with the Fieldpea crop, pea leaf miner, *Phytomyza horticola* Goureau, taxonomically described under the family Agromyzidae of the Order Diptera, is a pest of high economic importance (Spencer, 1973). This insect is highly polyphagous, with over 127 recognized host plants (Singh and Havi, 1982) recorded from 268 genera

from 36 families but most commonly on Brassicaceae, Fabaceae and Asteraceae (Spencer, 1990). It is one amongst the major insect pests of pea crop (Singh *et al.*, 1992) widely distributed over Africa, Asia and Europe (Crop Protection Compendium, 2007). The pea leaf miner is a serious and persistent pest of peas and Brassicas in northern India, wreaking havoc on these crops (Atwal *et al.*, 1969; Bhalla and Pawar, 1977; Prasad *et al.*, 1984). It can cause up to 90% damage to the crop by mining young leaves which results in stunting and low flower production (Tariq *et al.*, 1991). The tiny adults are two winged flies having greyish black mesonotum. The frons is yellow in colour. Eggs are laid by making punctures through inserting ovipositor into the leaf tissues. The adult females' activity, which punctures fragile leaves in several places with their sharp and pointed ovipositor for the purpose of oviposition or eating, is the first sign of damage to the leaves (Ahmad and Gupta, 1941). The damaging stages are larvae which are minute and slender and form a narrow, linear mine on the upper or lower leaf surface (Spencer, 1973) and feed on mesophyll between the upper and lower epidermis. Maggots that mine into the leaves, eating through the mesophyll while leaving the two epidermal layers intact, causes the most catastrophic harm to the crop (Ahmad and Gupta, 1941; Anceev and

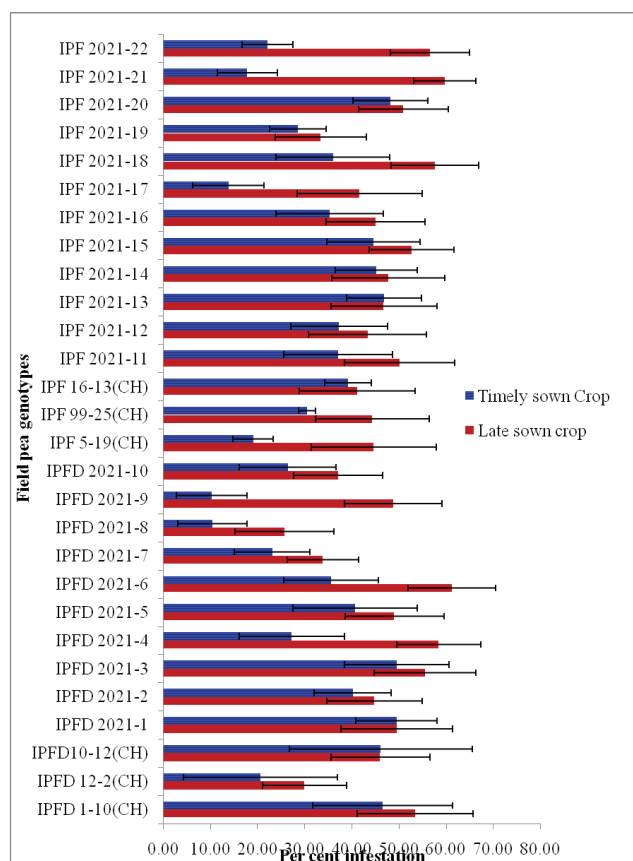


Fig 1. Per cent leaf infestation of different fieldpea genotypes caused by pea leaf miner in timely and late sown condition

Postolovski, 1978). Mining by larvae results in loss of chlorophyll which, in turn, affects the proper growth of plant and crop yield. Photosynthetic activity is severely hampered, and in severe infestations, leaves wither completely. The ability of afflicted plants to bloom and produce fruit is harmed. Under severe infestations, leaf miners cause withering of leaves and reduced flowering and fruiting (Molitas and Gabriel, 1978).

The majority of pest management methods are insecticidal in nature (Chopade, 1975; Dash, 1990; Khajuria and Sharma, 1995; Mehta *et al.*, 1995). However, due to their limits, insecticide spraying on vegetable crops has been severely restricted in recent years. As a result, there is a pressing need to investigate and implement new environmentally benign pest management strategies, such as the use of resistant/tolerant types to reduce the use of harmful chemicals. In the present scenario of preferring ecofriendly management avenues over chemical insecticides, development of varieties resistant to insect pest always gets the first thought. Deployment of host plant resistance in insect pest management strategies will render reduced losses, less insecticide use, better crop yields and safer environment (Howe and

Jander, 2008). Selections of varieties less prone to insect attack and studies related to adjustment in the date of sowing to gain escape resistance is the present day way forward (Chandra *et al.*, 2021). The mechanism of plant resistance rotates around biochemical host attributes too which affects the oviposition and feeding preferences of an insect pest. In view of this, the study was taken up to investigate how the fieldpea crop escapes leaf miner infestation by changing the time of its sowing and to see the correlation of infestation with some primary and secondary plant metabolites.

MATERIALS AND METHODS

Screening of fieldpea genotypes

A field experiment was conducted during rabi 2020-21 at the research farm of ICAR-Indian Institute of Pulses Research, Kanpur, Uttar Pradesh, India, to study the influence of sowing time on the level of infestation of pea leaf miner, *Phytomyza horticola* Goureaux. The site of experiment was geographically situated between 26.49188° N and 80.27748° E with an altitude of 128 m above mean sea level. A panel of twenty eight diverse field pea genotypes including checks was selected for the trial. Timely sowing was done on November 1, whereas another set for late sowing was done on December 9. The experiment was executed in randomized complete block design with three replications. In each replication, genotypes were planted in three rows of 4.0 m length with inter and intra row spacing of 60 cm and 10 cm, respectively. The crop was raised following recommended package of practices to maintain a normal healthy crop. No control measures adopted for insect pest control.

The incidence of leaf miner in field pea crop noticed from first week of February, at the time crop reached the flowering stage. For estimation, done at weekly interval, ten plants were tagged randomly in each genotype. The per cent leaf infestation was calculated by counting the infested leaves out of the total number of leaves present in the plant. On the basis of per cent leaf infestation, the genotypes were categorized using a scale of 1-9 ('1' being 'Very highly resistant' and '9' being 'Very highly susceptible'), following the method given by Singh and Weigand (1994).

Estimation of plant metabolites

Protein content, total sugar, total phenol and tannin content were quantified in randomly selected 10 different field pea genotypes. Protein present in sample was estimated using Lowry's method (Lowry *et al.*, 1951). The principle behind the Lowry method of determining protein concentrations lies in the reactivity of the peptide nitrogen[s] with the copper [II] ions under alkaline

Table 1. Categorization of fieldpea genotypes on the basis of degree of susceptibility against pea leaf miner during different planting time

S.N.	Field Pea Genotypes	susceptibility category	
		Timely sown	Late sown
1	IPFD 1-10(CH)	MS	S
2	IPFD 12-2(CH)	MR	MR
3	IPFD 10-12(CH)	MS	MS
4	IPFD 2021-1	MS	MS
5	IPFD 2021-2	I	MS
6	IPFD 2021-3	MS	S
7	IPFD 2021-4	MR	S
8	IPFD 2021-5	MS	MS
9	IPFD 2021-6	I	S
10	IPFD 2021-7	MR	I
11	IPFD 2021-8	HR	MR
12	IPFD 2021-9	HR	MS
13	IPFD 2021-10	MR	I
14	IPF 5-19(CH)	HR	MS
15	IPF 99-25(CH)	MR	MS
16	IPF 16-13(CH)	I	MS
17	IPF 2021-11	I	MS
18	IPF 2021-12	I	MS
19	IPF 2021-13	MS	MS
20	IPF 2021-14	MS	MS
21	IPF 2021-15	MS	S
22	IPF 2021-16	I	MS
23	IPF 2021-17	HR	MS
24	IPF 2021-18	I	S
25	IPF 2021-19	MR	I
26	IPF 2021-20	MS	MS
27	IPF 2021-21	HR	S
28	IPF 2021-22	MR	S

VHR = Very highly resistant; 2. HR = Highly resistant; 3.R = Resistant; 4.MR = Moderately resistant; 5.I = Intermediate; 6. MS = Moderately susceptible; 7. S = Susceptible; 8. HS = Highly susceptible; 9. VHS = Very highly susceptible

Table 2. Quantity of protein, total sugar, total phenol and tannin in different field pea genotypes

Field pea genotypes	Protein (%)	Total sugar(mg/g of Leaf sample)	Total phenol (mgGAE/g of leaf sample)	Tannin (mgTAE/g of leaf sample)
IPED 1-10(CH)	5.63 (13.71)	13.16 (3.76)	2.81 (1.95)	3.31 (2.07)
IPFD 2021-1	5.36 (13.38)	14.88 (3.98)	2.31 (1.82)	2.72 (1.92)
IPFD 2021-3	3.65 (10.99)	6.65 (2.76)	2.25 (1.80)	2.80 (1.94)
IPFD 2021-5	4.20 (11.81)	13.16 (3.76)	2.31 (1.81)	3.09 (2.02)
IPFD 2021-8	5.50 (13.54)	5.77 (2.60)	2.15 (1.77)	3.17 (2.04)
IPF 2021-11	5.28 (13.27)	19.31 (4.50)	3.23 (2.05)	3.61 (2.14)
IPF 2021-15	6.22 (14.42)	29.53 (5.52)	3.16 (2.04)	3.97 (2.22)
IPF 2021-16	5.38 (13.39)	31.59 (5.70)	2.89 (1.97)	3.65 (2.15)
IPF 2021-21	5.40 (13.43)	18.21 (4.38)	3.29 (2.07)	3.64 (2.15)
IPF 2021-22	4.92 (12.80)	17.71 (4.32)	2.80 (1.94)	3.45 (2.11)
C.D	0.563	0.118	0.030	0.029
SE(m)	0.188	0.039	0.010	0.010
SE(d)	0.266	0.056	0.014	0.014
CV	2.490	1.651	0.911	0.807

Figures under parenthesis, corresponding to protein values, are arcsine transformation of the original percent values; Figures under parenthesis, other than protein values, are square root transformation of the original values.

conditions and the subsequent reduction of the Folin-Ciocalteu phosphomolybdic phosphotungstic acid to heteropoly molybdenum blue by the copper-catalyzed oxidation of aromatic acids. The absorbance was taken at 660 nm in spectrophotometer. A calibration curve was plotted using BSA (Bovine serum albumin) as standard and the protein content in the sample was calculated in percentage using plotted standard curve of BSA. Total phenolic content of the sample extracts was determined using Folin-Ciocalteu reagent following the method of Singleton and Rossi (1965). The absorbance of the samples was measured at 650 nm against reagent blank in spectrophotometer. The Calibration curve was plotted using gallic acid as standard and phenols in the sample was calculated as gallic acid equivalents (mgGAE/g sample) using standard curve of gallic acid. Tannins in the sample was estimated using Folin-Denis method based on the principle that tannin like compounds reduce

Phosphomolybdic acid in alkaline solution to produce highly coloured blue solution, the intensity of which is proportional to the amount of tannins (Schanderl, 1970). The absorbance of the samples was taken at 700 nm in spectrophotometer. The calibration curve was plotted using tannic acid as standard and the content in the sample was calculated as tannic acid equivalents (mgTAE/g sample) using standard curve of tannic acid. The total sugar in the sample was estimated by the method of Dey (1990). The absorbance was recorded at 490 nm in spectrophotometer. The calibration curve was plotted by using glucose as standard. The total soluble sugar content in the sample was calculated in mg/g of leaf sample using standard. The chlorophyll content in all the 28 field pea genotypes was estimated using SPAD-502 and expressed in relative SPAD meter values which are proportional to the chlorophyll content present in the leaf (Ling *et al.*, 2011).

Table 3. Correlation matrix showing Pearson's correlation coefficient amongst metabolites and per cent leaf infestation in field pea genotypes

	Chlorophyll	Per cent Infestation (late sown genotypes)	Per cent Infestation (timely sown genotypes)	Protein	Total sugar	Total Phenol
% Infestation (late sown genotypes)	0.285 NS					
% Infestation (timely sown genotypes)	0.232 NS	0.127 NS				
Protein		-0.017 NS	-0.165 NS			
Total sugar		0.259 NS	0.097 NS	0.538 NS		
Phenol		<u>0.636*</u>	-0.121 NS	0.538 NS	0.703*	
Tannin		0.324 NS	-0.312 NS	0.632*	0.768**	0.864**

Correlation analysis

Correlation analysis was done by calculation of Pearson's correlation coefficient through the OPSTAT analytical software to see relation, if any, between per cent leaf infestation done by pea leaf miner with quantity of protein, total phenol, tannin, total sugar and chlorophyll.

RESULTS AND DISCUSSION

Screening of field pea genotypes

The per cent leaf infestation of 28 field pea genotypes and their comparison under timely and late sown conditions has been depicted in the Figure 1. In timely sown condition, the per cent leaf infestation ranged from 3.44 (IPFD 2021-9) to 49.44 (IPFD 2021-1 and IPFD 2021-3). Other genotypes observed with comparatively higher per cent leaf infestation in timely sown condition were IPF 2021-20 (48.19%), IPFD 1-10 (Check) (46.53%), IPFD 10-12 (Check) (46.11%), IPF 2021-13 (46.81%), IPF 2021-14 (45.14%) and IPF 2021-15 (44.58%). Range of leaf infestation was found increased (25.67 for IPFD 2021-8 to 61.17 for IPFD 2021-6) in case of late sown crops.

Kooner and Singh (1980) observed the per cent infestation of pea leaves by leaf miner which was ranged from 2.0 to 31.7 in different varieties whereas Kashyap *et al.* (1982) found 33.6, 20.3, and 22.9 per cent pea leaf

miner infestation on Bonneville, Arkel and Harabona-B cultivars of field pea, respectively. In the present study, IPFD 12-2 (Check), IPFD 2021-7, IPFD 2021-10 and IPFD 2021-19 were some of the genotypes observed with low per cent leaf infestation of 29.92, 33.83, 37.08 and 33.33, respectively. On the other hand, genotypes like IPFD 1-10 (Check), IPFD 2021-3, IPFD 2021-4, IPF 2021-11, IPF 2021-15, IPF 2021-18, IPF 2021-20, IPF 2021-21 and IPF 2021-22 were recorded with high per cent leaf infestation of 53.42, 55.50, 58.42, 50.08, 52.67, 57.58, 50.92, 59.75 and 56.58, respectively. Genotypes when sown under late condition, had recorded higher per cent leaf infestation as compared to timely sown genotypes. In both timely and late sown genotypes, it was observed that the per cent leaf infestation increased with progress of time.

In timely sown crops, mean per cent leaf infestation was recorded minimum on 8 February with only 5.96, followed by 14.98 on 15 February, 23.99 on 24 February and maximum of 41.48 on 1 March 2021. Similarly, in late sown condition, minimum per cent leaf infestation of 7.13 was observed on 3 February, which increased to 12.25 on 10 February, to 21.05 on 17 February, to 36.29 on 24 February and a maximum of 57.16 was recorded on 3 March 2021.

All the 28 field pea genotypes were rated using 1-9 scale and were categorized from Very highly resistant (1) to Very highly susceptible (9). In case of timely planting,

only 9 genotypes were categorized under ‘Moderately Susceptible’, 7 genotypes each under ‘Moderately Resistant’ and ‘Intermediate’ and 5 genotypes grouped under ‘Highly Resistant’ category (Table 1). None of the genotypes categorized as ‘Susceptible’ or beyond in case of timely planting. In late sown condition, among 28 genotypes, 8 genotypes grouped under ‘Susceptible’ category, 2 genotypes under ‘Moderately Resistant’, three genotypes under ‘Intermediate’, whereas 15 genotypes were categorized as ‘Moderately Susceptible’. None of the genotypes could be categorized as ‘Very Highly Resistant’, ‘Highly Resistant’, ‘Resistant’, ‘Highly Susceptible’ and ‘Very Highly Susceptible’ in case of late planting.

Kashyap *et al.* (1982) reported no substantial differences in resistance to this pest among the various potential pea cultivars. Mahobe and Narsinghani (1986) found none of the pea lines with desired level of resistance to this miner; however, they reported some degree of resistance in the cultivars JP-179, JP-854, JP-747, JP-169-1 and JP-Batri Brown. Also, Dash *et al.* (1988) found no cultivars resistant to pea leaf miner attack; however, “early wonder” showed considerable resistance, followed by Arkel. Bhat (1988) examined 20 lines of pea for resistance to *C. horticola*, *Lampides boeticus* L., and *Bruchus pisorum* L. in Kashmir and found that different cultivars had various levels of infestation, but no variety was totally resistant to this pest. Sharma and Sharma (1991) classified 8 pea cultivars (IP-3, JP-169, PG-2, S-143, Sel-35, Sel-93, Sel— 3487, VG-1) as resistant to pea leaf miner, 22 as moderately susceptible and 11 as highly susceptible in field trials conducted in Himachal Pradesh. Thakur and Patial (2019) screened ninety-two pea germplasm against pea leaf miner where six pea genotypes (DPP 25G, DPPLMR 41, JI 1766 (2), JP 179, LMR 100, S143) exhibited high resistance, nineteen as resistant and seventeen as moderately resistant to the pest. In the present study, only two genotypes i.e. IPFD 2021-8 and IPFD 12-2 could demonstrate good level of resistance in both late and timely planting. These identified genotypes after further validation can be used as donor in resistance breeding program.

Quantification of plant metabolites in field pea germplasm

The estimated values of protein, total sugar, total phenol and tannin in selected field pea genotypes are shown in Table 2.

The difference in the contents of these bio-chemicals among the genotypes studied was found statistically significant. The protein content was estimated highest in IPF 2021-15 with 6.22 per cent and lowest in IPFD 2021-3 with only 3.65 per cent. The total sugar was

observed maximum in IPF 2021-15 with 29.53 mg/g leaf sample while minimum quantity was observed in IPFD 2021-8 and IPFD 2021-3 with 5.77 and 6.65 mg/g of leaf sample, respectively. Sepehya *et al.* (2015) reported maximum sugar content of 4.21 mg/g in field pea. Minimum content of total phenol, 2.15 mg GAE/g leaf sample was estimated in IPFD 2021-8 to the maximum of 3.29 mg GAE/g leaf sample in IPF 2021-21. Wang *et al.* (1998) estimated total phenolics in field pea which differed significantly among cultivars, ranging from 162 mg/kg DM (dry matter) (CE, catechin equivalents) to 325 mg/kg DM (CE). Zia-ul-Haq *et al.* (2013) showed that the contents of total phenols, flavonoids, and condensed tannins in seeds of four Pakistani pea cultivars ranged between 0.84–0.99, 0.09–0.17, and 0.57–0.68 mg/g, respectively. Hegedusova *et al.* (2015) reported the total polyphenol contents of six garden pea varieties as 1179.995±28.081 mg GAE/kg as the highest value and the lowest value as 674.505± 26.541 mgGAE/kg. The tannin content was found minimum in IPFD 2021-1 with only 2.72 mgTAE/g of leaf sample while maximum tannin content was recorded in IPF 2021-15 with 3.97 mgTAE/g leaf sample. Overall, it was observed that IPFD 2021-15 was the genotype observed with maximum content of protein, total sugar, total phenol and tannin content, whereas, IPFD 2021-3 and IPFD 2021-8 was found low in all of these parameters.

The chlorophyll contents in all the 28 field pea genotypes were estimated and expressed in relative SPAD meter values which remain proportional to the chlorophyll content present in the leaf (Ling *et al.*, 2011). The SPAD meter values ranged from 26.63, 27.41 and 28.99 in IPF 16-13 (Check), IPF 99-25 (Check) and IPF 2021-11 to maximum of 51.02, 48.71 and 47.68 in IPFD 2021-3, IPFD 10-12 (Check) and IPFD 2021-4, respectively. Golawska *et al.* (2010) reported chlorophyll ranging from 36.57 to 39.82 (SPAD meter values) in *P. sativum*. The difference among genotypes in terms of chlorophyll content was found statistically significant.

Correlation between per cent infestation and physiological attributes

Correlation analysis were done among per cent infestation observed in timely sown crops, late sown crops, their protein content, total sugar, total phenol and tannin content in randomly selected 10 genotypes. Correlations of chlorophyll content of all 28 genotypes were analysed with the corresponding per cent leaf infestation in both timely and late sown conditions. Of all the correlations, only per cent infestation observed in late sown condition was found significantly and positively correlated with the total phenol content with a correlation coefficient of 0.64 (Table 3).

Also, Sharma and Aggarwal (1983) observed positive correlation between population of jassid, *Amrasca biguttula biguttula* Ishida, with free phenol and the tannin content of cotton. On the other hand, Woodhead *et al.*, (1980) found high phenolic acid concentrations linked to reduce eating by several grasshoppers and the plant hoppers. Dass and Odak (1987) reported total phenols to be adversely linked with pod fly (*Melanagromyza obtusa* Malloch) infestation in *Cajanus cajan*. Nomura and Itioka (2002) discovered that tannin inhibits cutworm larvae from growing and that the inhibitory impact was proportional to the amount of tannin consumed. According to Chandra (2012), total chlorophyll, carotenoid and sugar in maize germplasm were significantly and positively correlated with the Leaf Injury Rating (LIR) recorded with respect to *Chilo partellus*. Contrastingly, Singh (1984) reported that chlorophyll content of a genotype was not related to resistance or susceptibility in *Brassica* strains to pea leaf miner. In another study, Padmavathi *et al.* (2013) found leaf folder damage leading to 57% loss in chlorophyll content, a 23% drop in PS II activity and a 23% reduction in relative water content. Saheb *et al.* (2018) reported proteins with a significant negative correlation with incubation period and fifth instar larval duration, phenols with a significant positive correlation with fifth instar duration, reducing sugars showed positive correlation with incubation period, fifth instar duration and a negative significant correlation with post-oviposition period of leaf bud borer in groundnut. Saleem *et al.* (2019) observed significant negative correlation between *S. litura* damage and protein and phenol content while significant positive correlation was found with reducing and total sugar.

CONCLUSION

Although, there exist a large variation in results pertaining to the nature of correlation between plant biochemical and their impact on biology of insect pests; in the present study, physiological attributes like protein, total sugar, tannin, chlorophyll content found no significant correlation with per cent infestation in the field pea crop. The significant positive correlation observed between per cent leaf infestation in late sown condition and phenol content of the genotypes may also be seen as an induction or plant response to higher infestation of leaf miner in late sown crop. Secondary metabolites like phenol may prolong the life stages of the insect, facilitating prolonged feeding and thereby increase the level of infestation. Most of the genotypes which showed susceptibility under late sown condition had expressed more level of resistance when timely sowing was done.

ACKNOWLEDGMENT

The authors acknowledge the facilities provided by the Indian Council of Agricultural Research-Indian Institute of Pulses Research, Kanpur, Uttar Pradesh, India. The authors also acknowledge the support provided by the Rani Lakshmi Bai Central Agricultural University, Jhansi, Uttar Pradesh, India.

REFERENCES

- Ahmed, T. and Gupta, R. L. 1941. The pea leaf-miner, *Phytomyza atricornis* (Meigen), in India. *Indian Journal of Entomology*, 3(1): 37- 49.
- Ambrose, M. J. 1995. From Near East center of origin, the prized pea migrates throughout world. *Diversity-Arlington Then Washington*, 11:118-119.
- Ancev, E. and Postolovski, M. 1978. Contribution to knowledge of *Phytomyza horticola* Goureau (Diptera: Agromyzidae). *Zastita Bilja*, 29(3): 273-277.
- Arnoldi, A., Zanoni, C., Lammi, C. and Boschini, G. 2015. The role of grain legumes in the prevention of hypercholesterolemia and hypertension. *Critical Reviews in Plant Sciences*, 34 (1-3): 144-168.
- Atwal, A. S., Chaudhary, J. P. and Ramzan, M. 1969. Studies on the biology and control of pea leafminer, *Phytomyza atricornis* Meigen. (Agromyzidae: Diptera). *Punjab Agricultural University Journal Research*, 8 (1): 163-169.
- Bhalla, O. P. and Pawar, A. D. 1977. A survey study of insect and non-insect pests of economic importance in Himachal Pradesh. *A survey study of insect and non-insect pests of economic importance in Himachal Pradesh*. D. N. Road, Bombay, India: Tiku and Tiku, Kitab Mahal, 15-17.
- Bhat, A. A. 1988. Preliminary screening of some pea cultivars against insect pests in Kashmir. *Progressive Horticulture*, 20(1-2): 167-168.
- Burstin, J., Gallardo, K., Mir, R. R., Varshney, R. K. and Duc, G. 2011. 20 Improving Protein Content and Nutrition Quality. In *Biology and breeding of food legumes* Ed. Pratap, A., and Kumar, J. CABI, pp. 314. <https://doi.org/10.1079/9781845937669.0314>.
- Chandra, A. 2012. Study on secondary metabolite (DIMBOA) and some other primary metabolites (chlorophyll, protein, sugar) in maize germplasm and their relation to *Chilo partellus* (Swinhoe)

- susceptibility. PhD, Indian Agricultural Research Institute, New Delhi, India.
- Chandra, A., Sujayanand, G. K. and Kumar, R. 2021. Influence of Sowing Dates and Host Crops on Population Incidence of Whitefly, *Bemisia tabaci* (Gennadius) in Greengram and Blackgram. *National Academy Science Letters*, 44 (5): 389-391. <https://doi.org/10.1007/s40009-020-01033-8>.
- Chopade, H. M. 1975. Chemical control of some major pests of field pea, *Pisum sativum*. *Entomologists News*, 5 (2): 17.
- Crop Protection Compendium. 2007. Edition © CAB International, Wallingford, UK.
- Dahl, W. J., Foster, L. M. and Tyler, R. T. 2012. Review of the health benefits of peas (*Pisum sativum* L.). *British Journal of Nutrition*, 108 (S1): S3-S10. <https://doi.org/10.1017/S0007114512000852>.
- Dash, A. N. 1990. Evaluation of some insecticides for their efficacy against the pea leaf miner, *Phytomyza atricornis* (Meigen). *Indian Journal of Plant Protection*, 18(2): 295-297.
- Dash, A. N., Patnaik, N. C. and Nandi, A. 1988. Reaction of some garden pea cultivars to the pea leaf mine (*Phytomyza atricornis* Heigen) (Diptera: Agromyzidae) in Orissa. *Environmental Ecology*, 6 (2): 530-531.
- Dass, S. B. and Odak, S. C. 1987. Biochemical basis of resistance in pigeonpea pod walls to pod fly *Melanogromyza obtusa* Malloch (Diptera: Agromyzidae). *Crop improvement*, 14 (1): 64-68.
- Dey, P. M. 1990. Oligosaccharides. In *Methods in plant biochemistry*, Academic Press., 189-219. <https://doi.org/10.1016/B978-0-12-461012-5.50011-2>.
- Dixit, G. P., Parihar, A. K. and Gupta, S. 2014. Perspective for increasing field pea production in India. *Handbook on minor and imported pulses of India*. Commodityindia.com. 37-39.
- Duc, G., Blancard, S., Hénault, C., Petit, M. S., Bernicot, M. H., et al. 2010. Prominent cool season pulse crop and an important part of sustainable cropping system of agriculture durable in Bourgogne. *Innovations Agronomiques*, 11: 157-173.
- Golawska, S., Krzyzanowski, R. and Lukasik, I. 2010. Relationship between aphid infestation and chlorophyll content in Fabacea species. *Acta Biologica Cracoviensia. Series Botanica*, 52(2). <https://doi.org/10.2478/v10182-010-0026-4>.
- Gupta, S. and Parihar, A. K. 2015. Broadening the genetic base of pulse crops. *Pulses: Challenges & Opportunities*, 86.
- Hegedúsová, A., Mezeyová, I., Timoracká, M., Šlosár, M., Musilová, J. and Juriková, T., 2015. Total polyphenol content and antioxidant capacity changes in dependence on chosen garden pea varieties. *Potravinárstvo*, 9(1). <https://doi.org/10.1038/hortres.2017.17>.
- Howe, G. A. and Jander, G. 2008. Plant immunity to insect herbivores. *Annual review of plant biology*, 59 (1): 41-66. <https://doi.org/10.1146/annurev.arplant.59.032607.092825>.
- Kashyap, R. K., Bhanot, J. P. and Verma, A. N. 1982. Relative susceptibility of pea cultivars against leaf miner, *Melanogromyza conavistae* Greathead (Diptera: Agromyzidae). *Haryana Agricultural University Journal of Research*. 12(1): 128-129.
- Khajuria, D. R. and Sharma, J. P. 1995. Efficacy of insecticides in controlling pea leafminer (*Chromatomyia horticola*) on seed crop of pea (*Pisum sativum*). *Indian Journal of Agricultural Sciences*, 65(5): 381-384.
- Kooner, B. S. and Singh, H. 1980. Performance of Some Cultivars of Field-Peas Against Leaf-Miner (*Phytomyza Horticola* Gour.). *Indian Journal of Horticulture*, 37(1): 101-102.
- Ling, Q., Huang, W. and Paul, J. 2011. Use of a SPAD-502 meter to measure leaf chlorophyll concentration in *Arabidopsis thaliana*. *Photosynthesis Research* 107(2): 209-214. <https://doi.org/10.1007/s11120-010-9606-0>.
- Lowry, C. O., Rosebrough, N., Farr, A. and Randall, R. 1951. Protein measurement with the Folin phenol reagent. *Journal of Biological Chemistry*, 193(1): 265-275. [https://doi.org/10.1016/S0021-9258\(19\)52451-6](https://doi.org/10.1016/S0021-9258(19)52451-6).
- Mahobe, J. and Narsinghani, V. G. 1986. Screening of pea varieties against leaf miner (*Phytomyza atricornis* Meigen). *Vegetable Science*, 13(1): 387-388.
- Marles, M. S., Warkentin, T. D. and Bett, K. E. 2013. Genotypic abundance of carotenoids and

- polyphenolics in the hull of field pea (*Pisum sativum* L.). *Journal of the Science of Food and Agriculture*, 93(3): 463-470. <https://doi.org/10.1002/jsfa.5782>.
- Mehta, P. K., Vaidya, D. N. and Kashyap, N. P. 1995. Efficacy and economics of some insecticides against pea leaf miner, *Chromatomyia horticola* (Goureau) on pea. *Journal of Insect Science*, 8:116-117.
- Mitchell, D. C., Lawrence, F. R., Hartman, T. J. and Curran, J. M. 2009. Consumption of dry beans, peas, and lentils could improve diet quality in the US population. *Journal of the American dietetic association*, 109 (5): 909-913. <https://doi.org/10.1016/j.jada.2009.02.029>.
- Molitas, L. P. and Gabriel, B. P. 1978. Biology of the pea leaf miner, *Phytomyza horticola* G. (Diptera: Agromyzidae) in Philippines. *Philippine Entomologist*, 3: 89-108.
- Nemecek, T., Richthofen, J. S. V., Dubois, G., Casta, P., Charles, R., *et al.* 2008. Environmental impacts of introducing grain legumes into European crop rotations. *European Journal of Agronomy*, 28 (3): 380 - 393. <https://doi.org/10.1016/j.eja.2007.11.004>.
- Nomura, M. and Itioka, T. 2002. Effects of synthesized tannin on the growth and survival of a generalist herbivorous insect, the common cutworm, *Spodoptera litura* Fabricius (Lepidoptera: Noctuidae). *Applied Entomology and Zoology*, 37 (2): 285 - 289. <https://doi.org/10.1303/aez.2002.285>.
- Padmavathi, C., Katti, G., Padmakumari, A. P., Voleti, S. R. and Subba Rao, L. V. 2013. The effect of leaf folder *Cnaphalocrocis medinalis* (Guenée) [Lepidoptera: Pyralidae] injury on the plant physiology and yield loss in rice. *Journal of Applied Entomology*, 137(4): 249-256. <https://doi.org/10.1111/j.1439-0418.2012.01741.x>.
- Parihar, A. K., Bohra, A. and Dixit, G. P. 2016. Nutritional benefits of winter pulses with special emphasis on peas and rajmash. In *Biofortification of food crops* (pp. 61-71). Springer, New Delhi. pp.61-71. https://doi.org/10.1007/978-81-322-2716-8_6.
- Prasad, D., Singh, K. M., Katiyar, R. N. and Singh, R. N. 1984. Incidence of insect pests in early maturing high yielding variety of pea, *Pisum sativum* Linn. *Indian Journal of Entomology*, 46 (3): 353 - 362.
- Rubiales, D., Gonzalez - Bernal, M. J., Warkentin, T. D., Bueckert, R., Vaz Patto, M. C. *et al.* 2019. Advances in breeding of peas (Fabaceae). In *Achieving sustainable cultivation of vegetables* Ed. Hochmuth, G., Burleigh, Dodds Science Publishing. <https://doi.org/10.19103/AS.2019.0045.28>.
- Saheb, Y. P., Hari-Prasad, K. V., Swarajya, K. L. and Rani, J. S. 2018. Mechanisms of resistance in groundnut genotypes against leaf bud borer, *Anarsia ephippias* (Meyrick). *Journal of Entomology and Zoology Studies*, 6 (2): 694 - 705.
- Saleem, M. A., Gopalakrishna, N. K., Tippannavar, P. S. and Nadaf, H. L. 2019. Biophysical and biochemical mechanism of resistance to *Spodoptera litura* in groundnut (*Arachis hypogaea* L.). *Journal of Entomology and Zoology Studies*, 4: 86-96.
- Schanderl, S. H. 1970. Method in food analysis. New York: Academic Press, 709.
- Sepehya, S., Bhardwaj, S. K. and Dhiman, S. 2015. Quality Attributes of Garden Pea (*Pisum sativum* L.) as Influenced by Integrated Nutrient Management Under Mid Hill Conditions. *Journal of Krishi Vigyan*, 3 (2): 78 - 83. <https://doi.org/10.5958/2349-4433.2015.00017.3>.
- Sharma, H. C. and Agarwal, R. A. 1983. Role of some chemical components and leaf hairs in varietal resistance in cotton to jassid, *Amrasca biguttula biguttula* Ishida. *Journal of entomological research*, 7:145-149.
- Sharma, I. D. and Sharma, K. C. 1991. Screening of pea germplasm for resistance to leaf miner, *Phytomyza atricornis* Meigen. *Indian Journal of Plant Protection*, 19 (2): 206.
- Singh, P. 1984. Morphological, anatomical and biochemical basis of resistance in rapeseed and mustard to the leaf miner, *Phytomyza horticola* Goureau. PhD Thesis, Punjab Agricultural University, Ludhiana, Punjab, India.
- Singh, S. K., Rahman, S. J., Gupta, B. R. and Kalha, C. S. 1992. An integrated approach to the management of the major diseases and insect pests of peas in India. *International Journal of*

- Pest Management*, 38(3):265-267. <https://doi.org/10.1080/09670879209371706>.
- Singh, P., and Havi, G. S. 1982. A critical review on the distribution and host plant index of the leaf miner, *Phytomyza horticola* Goureau. *Bulletin of Pure and Applied Sciences*, 1: 45-51.
- Singh, K. B. and Weigand, S. 1994. Identification of resistant sources in *Cicer* species to *Liriomyza cicerina*. *Genetic Resources and Crop Evolution*, 41:75-79.
- Singleton, V. L. and Rossi, J. A. 1965. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *American journal of Enology and Viticulture*, 16 (3): 144-158.
- Spencer, K. A. 1973. Leaf miners on cereals and other graminaceous crops. In *Agromyzidae (Diptera) of Economic Importance*. Springer, Dordrecht, 236-297. https://doi.org/10.1007/978-94-017-0683-4_9.
- Spencer, K. A. 1990. Host Specialization in the World *Agromyzidae (Diptera)* with Illustrations by Ann Spencer. <https://doi.org/10.1007/978-94-009-1874-0>.
- Tariq, M., Khokhar, K. M., Farooq, M. and Mohammad, A. 1991. Larval fluctuation of pea leaf miner on pea crop and effect of abiotic factors on its dynamics. *Pakistan Journal of Agricultural Research*, 12(3): 202-205.
- Thakur, A. and Patiyal, M. 2019. Studies on host plant resistance in pea against pea leaf miner, *Chromatomyia horticola* (Goureau). *International Journal of Tropical Agriculture*, 37(2): 271-279.
- Wang, X., Warkentin, T. D., Briggs, C. J., Oomah, B. D., Campbell, C. G. and Woods, S. 1998. Total phenolics and condensed tannins in field pea (*Pisum sativum* L.) and grass pea (*Lathyrus sativus* L.). *Euphytica*, 101(1): 97-102.
- Woodhead, S., Padgham, D. E. and Bernays, E. A. 1980. Insect feeding on different sorghum cultivars in relation to cyanide and phenolic acid content. *Annals of Applied Biology*, 95(2):151-157. <https://doi.org/10.1111/j.1744-7348.1980.tb04733>.
- Zia-Ul-Haq, M., Amarowicz, R., Ahmad, S. and Riaz, M. 2013. Antioxidant potential of some pea (*Pisum sativum* L.) cultivars commonly consumed in Pakistan. *Oxidation Communications*, 36(4): 1046-1057.
- Zohary, D. and Hopf, M. 2000. Domestication of plants in the Old World: The origin and spread of cultivated plants in West Asia, Europe and the Nile Valley. Oxford university press.

MS Received: 17 November 2022

MS Accepted : 23 December 2022